

AD _____

Award Number: DAMD17-98-1-8168

TITLE: Phage Display Breast Carcinoma cDNA Libraries: Isolation of Clones Which Specifically Bind to Membrane Glycoproteins, Mucins, and Endothelial Cell Surface

PRINCIPAL INVESTIGATOR: Fumiichiro Yamamoto, Ph.D.

CONTRACTING ORGANIZATION: The Burnham Institute
La Jolla, California 92037

REPORT DATE: July 2001

TYPE OF REPORT: Final Addendum

PREPARED FOR: U.S. Army Medical Research and Materiel Command
Fort Detrick, Maryland 21702-5012

DISTRIBUTION STATEMENT: Approved for Public Release;
Distribution Unlimited

The views, opinions and/or findings contained in this report are those of the author(s) and should not be construed as an official Department of the Army position, policy or decision unless so designated by other documentation.

20020124 373

REPORT DOCUMENTATION PAGEForm Approved
OMB No. 074-0188

Public reporting burden for this collection of information is estimated to average 1 hour per response, including the time for reviewing instructions, searching existing data sources, gathering and maintaining the data needed, and completing and reviewing this collection of information. Send comments regarding this burden estimate or any other aspect of this collection of information, including suggestions for reducing this burden to Washington Headquarters Services, Directorate for Information Operations and Reports, 1215 Jefferson Davis Highway, Suite 1204, Arlington, VA 22202-4302, and to the Office of Management and Budget, Paperwork Reduction Project (0704-0188), Washington, DC 20503

1. AGENCY USE ONLY (Leave blank)		2. REPORT DATE July 2001	3. REPORT TYPE AND DATES COVERED Final Addendum (01 Jul 00 - 30 Jun 01)	
4. TITLE AND SUBTITLE Phage Display Breast Carcinoma cDNA Libraries: Isolation of Clones Which Specifically Bind to Membrane Glycoproteins, Mucins, and Endothelial Cell Surface			5. FUNDING NUMBERS DAMD17-98-1-8168	
6. AUTHOR(S) Fumichiro Yamamoto, Ph.D.				
7. PERFORMING ORGANIZATION NAME(S) AND ADDRESS(ES) The Burnham Institute La Jolla, California 92037 E-Mail: fyamamoto@burnham.org			8. PERFORMING ORGANIZATION REPORT NUMBER	
9. SPONSORING / MONITORING AGENCY NAME(S) AND ADDRESS(ES) U.S. Army Medical Research and Materiel Command Fort Detrick, Maryland 21702-5012			10. SPONSORING / MONITORING AGENCY REPORT NUMBER	
11. SUPPLEMENTARY NOTES				
12a. DISTRIBUTION / AVAILABILITY STATEMENT Approved for Public Release; Distribution Unlimited				12b. DISTRIBUTION CODE
13. ABSTRACT (Maximum 200 Words) We attempted to identify phage display cDNA clones that exhibited proteins with carbohydrate affinity, using libraries constructed in mid copy display T7Select 10-3 vector. Although we hoped that higher affinity with multiple number of capsid fusions would facilitate the screening of those clones, the attempts have been unsuccessful.				
14. SUBJECT TERMS Breast cancer, Phage display libraries, cDNA cloning, Surface Binding, carbohydrate-binding proteins (lectins)				15. NUMBER OF PAGES 5
				16. PRICE CODE
17. SECURITY CLASSIFICATION OF REPORT Unclassified	18. SECURITY CLASSIFICATION OF THIS PAGE Unclassified	19. SECURITY CLASSIFICATION OF ABSTRACT Unclassified	20. LIMITATION OF ABSTRACT Unlimited	

Table of Contents

Cover.....	1
SF 298.....	2
Table of Contents.....	3
Introduction.....	4
Body.....	4
Key Research Accomplishments.....	4
Reportable Outcomes.....	5
Conclusions.....	5
References.....	5
Appendices.....	

Introduction:

In multicellular organisms, each cell is surrounded by other cells as well as a complex network of extracellular matrix. On the outer membrane, complex carbohydrate structures are present as parts of glycoproteins and glycolipids. These carbohydrate structures play an important role in interacting with proteins (lectins) (1) and other carbohydrate structures (2).

In an attempt to identify and clone unknown human proteins with carbohydrate affinity, we proposed to employ the novel technology named phage display (3). The phage display technology is based on the surface expression of the peptide sequences fused with phage capsid protein, and has been most successfully used in cloning phage particles that express variable domains of antibodies specific to certain antigens (4,5). Phage display peptide libraries made with synthetic oligonucleotides have also been utilized to identify peptide sequences that interact with a variety of bait ligands, such as proteins, peptides, DNAs, RNAs, and oligonucleotides (6,7). We have used the T7 phage cDNA display system developed by Novagen (Madison, WI) (8). Different from filamentous phage systems where the peptide sequences are fused with capsid proteins at the C-terminus, the T7 system allows the fusion of protein sequences up to 1200 amino acid residues long fused with gene 10 capsid protein at the N-terminus of proteins.

We constructed a phage display cDNA library using RNA from cells that stably expressed A transferase, and performed biopanning experiments using, as a bait ligand, crude mucin fraction containing blood group H-specific glycoproteins. Although no enrichment of the phages that expressed A transferase fusion protein was observed, selective augmentation was observed of the phages that expressed the fusion proteins with galectin-3, a soluble β -galactoside-binding (S-type) lectin (9). Because of this lectin's known affinity with the blood group-specific oligosaccharides (10,11), the results demonstrated that the phage display was useful in cloning cDNAs encoding a protein with binding capacity to carbohydrates.

Body:

During the originally proposed two years, we tried, without success, to identify cDNA clones that encode unknown proteins with carbohydrate affinity using the T7 phage system. The cDNA display libraries constructed in the T7Select 1-1 vector were primarily used in the screenings. Since the T7Select 1-1 vector displays a low copy number (0.1-1 capsid fusion per phage) of peptides or larger proteins, there was a possibility that the affinity was too weak with this system for the detection of unknown carbohydrate-binding protein(s). Because the new vector, T7Select 10-3 vector, was developed for mid copy number display (5-15 capsid fusions per phage) of peptides and proteins, and has become available, we have repeated some of the screening experiments using the libraries constructed in this mid copy vector in the no-cost extension period. Although we hoped to identify candidate novel lectin(s) using this T7Select 10-3 vector with higher affinity, no promising candidates have been obtained.

Key Research Accomplishments:

None

Reportable Outcomes:

None

Conclusions:

Although we re-tried to identify phage clones that express fusion proteins with affinity to carbohydrate ligands using mid copy display vector, the attempts have been unsuccessful. The reason for our failure is unclear. However, different from galectins, the lectins of other family may lose the carbohydrate binding affinity by the fusion at the N-terminus. It is possible that the expression of those fusion proteins may be toxic to bacteria and the phages that express those proteins may be eliminated from the population in the library. It is also possible that those proteins are instable and may not be displayed on phage particles.

References:

1. Drickamer, K. Molecular structure of animal lectins. in *Molecular Glycobiology* (eds. Fukuda, M. & Hinds, G.) 53-87 (IRL Press, Oxford, UK, 1994).
2. Kojima, N. & Hakomori, S. Specific interaction between gangliosylceramide (Gg3) and sialosylceramide (GM3) as a basis for specific cellular recognition between lymphoma and melanoma cells. *J Biol Chem* 264, 20159-20162. (1989).
3. Smith, G.P. Filamentous fusion phage: novel expression vectors that display cloned antigens on the virion surface. *Science* 228, 1315-1317 (1985).
4. Winter, G. & Milstein, C. Man-made antibodies. *Nature* 349, 293-299 (1991).
5. Clackson, T., Hoogenboom, H.R., Griffiths, A.D. & Winter, G. Making antibody fragments using phage display libraries. *Nature* 352, 624-628 (1991).
6. Wrighton, N.C. et al. Small peptides as potent mimetics of the protein hormone erythropoietin [see comments]. *Science* 273, 458-464 (1996).
7. *Phage Display of Peptides and Proteins: A Laboratory Manual*, (Academic Press, San Diego, 1996).
8. Dunn, J.J. & Studier, F.W. Complete nucleotide sequence of bacteriophage T7 DNA and the locations of T7 genetic elements. *J Mol Biol* 166, 477-535. (1983).
9. Yamamoto, M., Kominato, Y. & Yamamoto, F. Phage display cDNA cloning of protein with carbohydrate affinity. *Biochem Biophys Res Commun* 255, 194-199 (1999).
10. Abbott, W.M., Hounsell, E.F. & Feizi, T. Further studies of oligosaccharide recognition by the soluble 13 kDa lectin of bovine heart muscle. Ability to accommodate the blood-group-H and -B-related sequences. *Biochem J* 252, 283-287 (1988).
11. Sato, S. & Hughes, R.C. Binding specificity of a baby hamster kidney lectin for H type I and II chains, poly-lactosamine glycans, and appropriately glycosylated forms of laminin and fibronectin. *J Biol Chem* 267, 6983-6990 (1992).